

USE OF TIGECYCLINE (TYGACIL) AGAINST DRUG RESISTANT ORGANISMS IN EASTERN REGION

¹MUGHIS UDDIN AHMED, ²JAVED MEMON, ³SHABAN MOHAMMED, ⁴NEARCHOS YIANNAKOU
⁵RIFAT REHMANI, ⁶NADIA AL-HAWASHIM AND ⁷MOTASIM AHMED

¹Department of Pathology and Laboratory Medicine, King Abdulaziz Hospital (NGHA) Al-Ahsa KSA
²ICU King Abdulaziz Hospital (NGHA) Al-Ahsa KSA. ³⁻⁵King Abdulaziz Hospital (NGHA) Al-Ahsa KSA
⁶Pathology Eastern Region. ⁷Al-Moosa General Hospital, Al-Ahsa KSA

ABSTRACT

Objective:

This study was designed to look for sensitivity and resistant pattern of tigecycline in different gram positive and gram negative infections which were resistant to different antibiotics and also look for various methods to prevent drug resistance against tigecycline (tygacil) group of antibiotic.

Materials and Methods:

Three hundred seventy five (375) isolates which includes MRSA (Methicillin Resistant Staphylococcus Aureus), VRE (Vancomycin Resistant Enterococci), ESBL (Extended Spectrum Beta Lactamase), *Stenotrophomonas maltophilia* and MDR (Multi Drugs Resistant) *Acinetobacter* species were identified with the help of colonial characteristics, gram staining, biochemical reactions including API strips system, and special techniques used for each organism. Sensitivity was done with help of disc diffusion (Kirby Bauer) method for tigecycline (tygacil) 15 ug disc provided by company.

Results:

This is a retrospective study which has showed that MRSA were 100% sensitive to tigecycline and VRE were also 100% sensitive to this antibiotic. The ESBL were 90% sensitive and *Stenotrophomonas maltophilia* 87% to tigecycline. The MDR *Acinetobacter* species were only 41% resistant which was high in 2008 as compared to overall sensitivity pattern. Male and female were almost equal in this study. Highest number of cases was reported from 70-80 years age group. The different isolates were from different locations from hu-

man body and different wards including ICU (Intensive Care Unit).

Conclusions:

Tigecycline exhibit high in vitro activity against most of the commonly encountered gram positive and gram negative resistant organisms which were pathogens in this region. We should take care not to get antibiotic resistance to be developed against tigecycline by appropriate uses and preventive measures (hand hygiene etc.)

Keywords:

Tigecycline (Tygacil), MRSA, VRE, ESBL, Antibiotic Resistance

INTRODUCTION

We come across different type of organisms including gram positives, gram negatives, atypicals, anaerobes, resistant gram positives and resistant gram negatives. Many organisms are sensitive to different antibiotics but some organisms like resistant

Gram positive (MRSA) and resistant gram negatives (ESBL) are problem to treat¹⁻⁸. The new antibiotic by the name of tigecycline (tygacil) has very good in vitro activity against resistant gram positives and gram negatives besides other organisms mentioned earlier⁹⁻¹³. This antibiotic belongs to the family of glycoycline which is semi-synthetic derivatives of the tetracycline. Tigecycline is the (tigecycline 9-t-butylglycylamide-minocycline) first compound in the new glycoycline class of antimicrobials to become available for clinical use^{1,2,10,14,17}. A lot of work has been done on this

antibiotic in North America, Europe and other countries. One study was done on this antibiotic in the Beirut- Lebanon from Middle East. No published data on this antibiotic available from the Saudi Arabia¹⁸⁻²⁷.

As there is no data available locally. This study will provide a guide on product use of this antibiotic in this part of the world. It will also provide guideline to physicians and surgeons. It will also give information about the proper usage of this product in certain reserve situations like resistant infections. We should concentrate to stop misuse, disuse, overuse and abuse of this antibiotic among hospitalized patients as well as out-patients. This antibiotic should be kept as reserve medicine to be used in resistant cases where no other antibiotic is available²⁵⁻²⁷.

MATERIALS AND METHODS

Three hundred seventy five (375) isolates which includes MRSA (Methicillin Resistant *Staphylococcus Aureus*), VRE (Vancomycin Resistant *Enterococcus*), ESBL (Extended Spectrum Beta Lactamase), *Stenotrophomonas maltophilia* and MDR (Multi Drugs Resistant) *Acinetobacter* species were identified with the help of colonial characteristics, gram staining, biochemical reactions including API strips system, and special techniques used for each organism. Sensitivity was done with help of disc diffusion (Kirby Bauer) method for tigecycline (tygacil) 15 ug disc provided by company. API strips used were 20 E, NE, Staph.API, Vancomycin resistant identification disc & Etest. The other identifications disc like methicillin or cloxacillin, ESBL identifications disc, other biochemical reactions and specialized techniques for identification were also used.

RESULTS

This antibiotic has been used in 375 different organism isolated from January 2008-December 2010 from our hospital. In this is a retrospective study in which tigecycline was used on MRSA, ESBL, VRE, MDR *Acinetobacter* species and

Stenotrophomonas maltophilia. This study showed that MRSA were 97 (100%) sensitive to tigecycline and VRE were also 07(100%) sensitive to this antibiotic. The ESBL were 173 (90%) sensitive and 20 (10%) were resistant to tigecycline. The *Stenotrophomonas maltophilia* 13 (87%) sensitive and 02 (13%) resistant to tigecycline. The MDR *Acinetobacter* species were only 26 (41%) sensitive, which was high in 2008 as compared to overall sensitivity pattern and 37 (59%) resistant to tigecycline (Table 1 & 2). ESBLs were *Escherichia coli*, *Klebsiella pneumoniae*, *Klebsiella oxytoca* and *Proteus mirabilis* in different numbers and percentages. In this highest was *Escherichia coli* 144(74.6%) with sensitivity of 94.4%, followed by *Klebsiella pneumoniae* 40 (20.7%) with 82.5% sensitivity, *Proteus mirabilis* 08 (4.2%) with sensitivity around 62.5% and *Klebsiella oxytoca* which was 0.5% amongst organisms with sensitivity 100% (Table 3). Male and female were almost equal in this study. Highest number of cases were reported from 70-80 years age group (Table 4). The different isolated were from different locations from human body and different wards including (Intensive Care Unit) ICU (Table 5).

DISCUSSION

Our study has shown that tigecycline in vitro very good activity against different antibiotic resistance organisms like MRSA, VRE, ESBL and *Stenotrophomonas maltophilia* but it showed less activity against MDR *Acinetobacter* species, which was less in year 2008 and increased by the end of year 2010. We should look for factors responsible for this resistance. Different studies provided us tigecycline activity against different organisms. In USA sepsis caused by *Elizabethkingia miricola* successfully treated with tigecycline²⁹. Bacterial isolates were consecutively collected between (2004-2007) from 24 countries of European Union, tigecycline was highly active in vitro against most of the pathogens monitored, including MDR pathogens. This activity was stable over the study period, which increasing resistance was noted for several comparator agents during the same period. Tigecycline is approvmentarium used for the treatment of intra-abdominal and complicated Skin and soft tissue

TABLE NO.1
DIFFERENT ORGANISMS WITH SENSITIVITY AND RESISTANCE PATTERN FOR TIGECYCLINE

| Name of organisms | Number of Sensitive organisms | Percentage of Sensitive organisms | Number of Resistant organisms | Percentage of Resistant organisms | Total Number of organisms | Percentage of Total Organisms |
|----------------------------|-------------------------------|-----------------------------------|-------------------------------|-----------------------------------|---------------------------|-------------------------------|
| MRSA | 97 | 100% | 0 | 0% | 97 | 100% |
| VRE | 7 | 100% | 0 | 0% | 7 | 100% |
| ESBL | 173 | 90.00% | 20 | 10.00% | 193 | 100% |
| Stenotrphomonas maltophila | 13 | 87.00% | 2 | 13.00% | 15 | 100% |
| MDR Acinetobacter spp. | 26 | 41.00% | 37 | 59.00% | 63 | 100% |
| Total | 316 | 84.00% | 59 | 16.00% | 375 | 100% |

TABLE NO.2
SEX DISTRIBUTION

| Group | Male | Percentage | Female | Percentage | Total | Percentage |
|----------------------------|------|------------|--------|------------|-------|------------|
| MRSA | 54 | 56.00% | 43 | 44.00% | 97 | 100.00% |
| VRE | 2 | 29.00% | 5 | 71.00% | 7 | 100.00% |
| ESBL | 87 | 45.00% | 106 | 55.00% | 193 | 100.00% |
| Stenotrphomonas maltophila | 12 | 80.00% | 3 | 20.00% | 15 | 100.00% |
| MDR Acinetobacter spp. | 30 | 48.00% | 33 | 52.00% | 63 | 100.00% |
| Total=193(Male+Female) | 185 | 49.00% | 190 | 51.00% | 375 | 100.00% |

TABLE NO.3
ESBL ORGANISMS ISOLATED

| Name of ESBL isolated | Number of isolated | Percentage of isolates | Number of Sensitive | Percentage of Sensitive | Number of Resistant | Percentage of Resistant |
|-----------------------|--------------------|------------------------|---------------------|-------------------------|---------------------|-------------------------|
| Escherichia coli | 144 | 74.60% | 136 | 94.4% | 8 | 5.60% |
| Klebseilla pneumoniae | 40 | 20.70% | 33 | 82.50% | 7 | 17.50% |
| Klebseilla oxytoca | 1 | 0.50% | 1 | 100% | 0 | 0.00% |
| Proteus mirabilis | 8 | 4.20% | 5 | 62.50% | 3 | 37.50% |
| Total | 193 | 100% | 175 | 91.00% | 18 | 9.00% |

TABLE NO.4
VARIOUS SITE FROM BODY ORGANISMS ISOLATED

| Site of isolation | MRSA | VRE | ESBL | Stenotrophomonas maltophilia | MDR Acinetobacter spp. |
|-------------------|-----------|----------|------------|------------------------------|------------------------|
| Nose | 35 | 0 | 0 | 0 | |
| Wound | 18 | 1 | 29 | 2 | 25 |
| Trachea | 2 | 0 | 3 | 0 | 13 |
| Endo | 1 | 0 | 3 | 4 | 3 |
| Abscess | 17 | 0 | 5 | 1 | 1 |
| Sputum | 4 | 0 | 2 | 3 | 5 |
| Tissue | 2 | 0 | 6 | 0 | 2 |
| Drain | 2 | 0 | 3 | 0 | 1 |
| Throat | 1 | 0 | 0 | 0 | 0 |
| Blood | 3 | 5 | 14 | 3 | 2 |
| Bed sore | 1 | 0 | 0 | 0 | 0 |
| Ear | 1 | 0 | 0 | 0 | 0 |
| Axilla | 1 | 0 | 0 | 0 | 0 |
| Hand | 1 | 0 | 0 | 0 | 0 |
| Abdomen | 1 | 0 | 0 | 0 | 0 |
| Synvial fluid | 1 | 0 | 0 | 0 | 0 |
| Urine | 0 | 1 | 121 | 0 | 8 |
| Tip | 0 | 0 | 1 | 2 | 1 |
| BF | 0 | 0 | 1 | 0 | 0 |
| Eye | 0 | 0 | 5 | 0 | 1 |
| NPA | 0 | 0 | 0 | 0 | 1 |
| Total | 97 | 7 | 193 | 15 | 63 |

TABLE NO.5
DIFFERENT AGE GROUP FROM WHERE ORGANISMS ISOLATED

| Age Group | MRSA | VRE | ESBL | Stenotrophomonas maltophilia | MDR Acinetobacter spp. |
|---------------|-----------|----------|------------|------------------------------|------------------------|
| 0-10 years | 15 | 0 | 7 | 3 | 0 |
| 10-20 years | 2 | 0 | 11 | 1 | 5 |
| 20-30 years | 9 | 0 | 14 | 0 | 4 |
| 30-40 years | 6 | 0 | 18 | 0 | 6 |
| 40-50 years | 7 | 2 | 22 | 0 | 10 |
| 50-60 years | 9 | 0 | 23 | 2 | 6 |
| 60-70 years | 20 | 0 | 27 | 1 | 16 |
| 70-80 years | 22 | 4 | 31 | 5 | 11 |
| 80-90 years | 7 | 1 | 35 | 1 | 3 |
| 90-100 years | 4 | 0 | 3 | 2 | 1 |
| 100-110 years | 0 | 0 | 2 | 0 | 1 |
| Total | 97 | 7 | 193 | 15 | 63 |

infections and retain potential to be a valuable part of the armamentarium used to combat MDR pathogen in Europe¹².

Tigecycline exhibits potent activity against *Acinetobacter* species in USA comparable to that shown against species of *Enterobacteriaceae* 2,8,9,13,22,23. Resistance of this pathogen (*Streptococcus pneumoniae*) is prevalent and growing local resistance profiles should be consulted before selecting empiric therapy in USA¹⁹. Complicated Skin & Skin-structure infections, intra-abdominal infections, surgical site infections and surgical infections MDR infections can be controlled by proper and adequate treatment. Prevention of infections can be done by adopting hand hygiene, other preventive and antibiotic formulary resistance program (antibiotic stewardship) in USA¹⁴. Tigecycline appeared safe and efficacious in patients with selected serious infections due to resistant gram-negative organisms including *Enterobacter* species, *Acinetobacter baumannii* and *Klebsiella pneumoniae* in USA¹⁵.

Tigecycline exhibits high in vitro activity against most of the commonly encountered ESBL producing and MDR bacterial pathogens in Beirut, Lebanon Middle East¹⁶. There is significant data in favor of the clinical use of tigecycline in management of different infections in UK. Tigecycline satisfy the need for a new broad-spectrum antibiotic class from Latin America. The in vitro activity of Tigecycline against both gram-positive and gram negative isolates indicates that it may be a useful for the treatment of nosocomial infections caused by organisms resistant to other antibacterial agents¹⁸.

Tigecycline demonstrated excellent activity against clinically resistant organisms in USA¹⁹. High tissue penetration into the epithelial lining fluid of infected and un-infected murine lungs by tigecycline in USA. Minimum Inhibitory Concentration (Mic.) of tigecycline is less than 2(.16) is sensitive and 4(13-15) is intermediate and more than >8(<12) is resistant for *Acinetobacter* species¹⁰⁻¹¹.

CONCLUSION

From this study we can draw following conclusion that gram positive and gram negative antibiotic resistance infections are quite prevalent in our region and tigecycline is new antibiotic which can be used in these resistant infections. We should also make proper guidelines for proper use of this anti-

otic to avoid antibiotic resistance against this new class of semi-synthetic antibiotic. This antibiotic use is by proper healthcare professional permission who can understand gravity of antibiotic resistance. To prevent drug resistance antibiotic stewardship and preventive measures like hand hygiene are necessary. This magic bullet can be used for long period without problem of resistance.

RECOMMENDATIONS

It needs wider studies in different region of Saudi Arabia and a committee should be made to draw policies and guideline for antibiotics used in general and tigecycline in particular to overcome the problem of antibiotics resistance.

REFERENCES

1. Chopra I. Glycylines: third-generation tetracycline antibiotics. *Current Opinion in Pharmacology*.2001;1:464-469:
2. Bradford P. A first in class glycycline. *Clin Micro Newsl* 2004;26:163-168
3. Chopra I and Robert M. Tetracycline antibiotics: Mode of action, Application, Molecular Biology and Epidemiology of Bacterial Resistance. *Microbiol. Mol. Biol. Rev.* 2001;65:232-260
4. Bauer G., Berens C., Projan SJ et al. Comparison of tetracycline and tigecycline binding to ribosomes mapped by dimethylsulphate and drug-directed Fe²⁺ cleavage of 16S rRNA. *J Antimicrob Chemother* 2004;53:592-599
5. Olson MW., Ruzin A., Feyfant E et al. Functional, biophysical and structural bases for antibacterial activity of tigecycline. *Antimicrob Agents Chemother* 2006;50:2156-2166
6. Zhanel GG., Homenuik K., Nichol K et al. The glucycline: a comparative review with the tetracyclines. *Drugs* 2004;64:63-88
7. Keeney D., Ruzin A., Bradford PA and Ram A. A transcriptional regulator, and AcrAB, an RND-type efflux pump, are associated with decreased susceptibility to tigecycline in *Enterobacter cloacae*. *Microb Drug resist* 2007;13:1-6
8. Ruzin A., Keeney D., and Bradford PA. AdeABC multidrug efflux pump is associated with decreased susceptibility to tigecycline in *Acinetobacter calcoaceticus*-*Acinetobacter baumannii* complex. *J Antimicrob Chemother* 2007;59:1001-1004
9. McAleese F., Petersen P., Ruzin A et al. A novel MATE family efflux pump contributes to the reduced susceptibility of laboratory-derived *Staphylococcus aureus* mutants to tigecycline. *Antimicrob Agents Chemother* 2005;49:1865-1871

10. Wyeth Pharmaceutical Inc Tygacil package insert 2005
11. Sultan M. Letter to Doctor from Medical Adviser Wyeth Pharmaceutical Inc Tygacil letters to doctors 2005
12. Norskov-Lauritsen N., Marchandin H., and Dowzicky MJ. Antimicrobial susceptibility of tigecycline and comparators against bacterial isolates collected as part of the TEST study in Europe(2004-2007). *Int J Antimicrob Agents* 2009;34:121-130
13. Jones RN., Ferraro MJ., Reller LB et al. Multicenter Studies of Tigecycline Disk Diffusion Susceptibility Result for *Acinetobacter* spp. *J Clin Microbiol* 2007;45(1):227-230
14. Evans HL., and Sawyer RG. Preventing Bacterial Resistance in Surgical patients. *Surg Clin N Am* 2009;89:501-518
15. Vasilev K., Reshedko G., Orasan R et al. A phase 3, open-label, non-comparative study of tigecycline in the treatment of patients with selected serious infections due to resistant Gram-negative organisms including *Enterobacter baumannii* and *Klebsiella pneumoniae*. *J Antimicrob Chemother* 2008;62 Suppl. 1:i29-i40
16. Araj GF., and Ibrahim GY. Tigecycline in vitro activity against commonly encountered multidrug-resistant Gram-negative pathogens in a Middle Eastern country. *Diagnostic Microbiology and Infectious Disease* 2008;62:411-415
17. MacGowan AP. Tigecycline pharmacokinetic/pharmacodynamic update. *J Antimicrob Chemother* 2008;62 suppl. 1:i11-i16
18. Wilcx MH. Tigecycline and the Need for a New Broad-Spectrum Antibiotic Class. *Surgical Infections* 2006;7(1):69-80 Rossi F., Garcia P., Ronzon B et al. Rates of Antimicrobial Resistance in Latin America (2004-2007) and vitro Activity of the Glycycline Tigecycline and Other Antibiotics. *BJID* October 2008; 12(5):405-415
19. Low DE., Markovic., and Dowzicky MJ. Antimicrobial Susceptibility Among Bacterial Isolates from ICU and Non-ICU Settings and Different Age Groups: Results from the Tigecycline Evaluation and Surveillance Trial in North America. *Journal of Chemotherapy* 2009;21(1):16-23
20. Crandon JL., Kim A., and Nicolau DP. Comparison of tigecycline penetration into the epithelial lining fluid of infected and uninfected murine lungs. *J Antimicrob Chemother* August 2009;10:301-303
21. Petersen PJ. and Bradford PA. Effect of medium age and supplementation with the biocatalytic oxygen-reducing reagent oxyrase on in vitro activities of tigecycline against recent clinical isolates. *Antimicrob Agents Chemther* 2005;39:3910-3918
22. Fritsche TR., Sader HS., Stilwell MG et al. Potency and spectrum of tigecycline tested against an international collection of bacterial pathogens associated with skin and soft tissue infections. *Diagn Microbiol Infect Dis* 2005;52:195-201
23. Sader HS., Jones RN., Dowzicky MJ et al. Antimicrobial activity of tigecycline tested against nosocomial bacterial pathogens from patients hospitalized in intensive care unit. *Diagn Microbiol Infect Dis* 2005;52:203-208
24. Brown SD and Tracewski MM. Comparative in vitro antimicrobial activity of tigecycline a new compound in freshly prepared medium and quality control. *J Clin Microbiol* 2007;45:2173-2179
25. Bradford PA., Weaver-Sands DT and Petersen PJ. In vitro activity of tigecycline against isolates from patients enrolled in phase 3 clinical trials of treatment for complicated skin and skin-structure infections and complicated intra-abdominal infections. *Clin Infect Dis* 2005;41 Suppl5:S315-S332
26. Luna VA., King DS., Gullledge J et al. Susceptibility of *Bacillus anthracis*, *Bacillus cereus*, *Bacillus mycoides*, *Bacillus pseudomycoides* and *Bacillus thurriensis* to 24 antimicrobials using Sensititre automated microbroth dilution and Etest agar gradient diffusion methods. *J Antimicrob Chemother* 2007;60:555-567
27. Fritsche Tr., Strabala PA., Sader HS et al. Activity of tigecycline tested against a global collection of Enterobacteriaceae, including tetracycline-resistant isolates. *Diagn Microbiol Infect Dis* 2005;52:209-213
28. Jacobus NV., McDermott LA., Ruthazer R et al. In vitro activities of tigecyclines against the *Bacteriodes fragilis* group. *Antimicrob Agents Chemother* 2004;48:1034-1036
29. Green ON., Murray P., and Gea-Banacloche JC. Sepsis caused by *Elizabethkingia miricola* successfully treated with tigecycline and levofloxacin. *Diagn Micro Infect Dis* 2008;62:430-432.